

T2-weighted MRI of breast cancer using hyaluronic acid coated magnetic nanoparticles as a tool of contrast agent

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ABSTRACT Novel diagnostic technique has been developed in many research area using targetable contrast agents with magnetic resonance imaging (MRI) for cancer diagnosis. It is efficient for cancer diagnosis to use MRI with biocompatible targeting moiety and magnetic nanoparticles (MNPs). Thus, we synthesized MNPs using thermal decomposition method which enable sensitive T2- or T2 Turbo spin echo (TSE) weighted magnetic resonance imaging.

And it was coated with Hyaluronic acid (HA). Also we carried out that breast cancer cell line (MDA-MB-231) which has cancer stem cell property was injected in heterotopic mouse model. And then magnetic resonance sequence (T2) for imaging effects and targeting ability were analyzed into MNPs conjugated HA. We noted that MDA-MB-231 cell which high-expressed CD44 ligand was showed contrast enhance efficiency through magnetic nanoparticles because of combining a lot of HA. As a result of these studies, we conclude that HA coated magnetic nanoparticles can be effectively used as a novel probe for visualizing of Breast cancer stem cell.

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1. Introduction

Molecular imaging provides as a tool to diagnose cancer at the cellular and molecular levels. It not only allows early and accurate tumor localization in diagnostic cancer imaging, but also has a potential to visualize the biological processes of tumor growth, metastasis and response to treatment.(1-10) Molecular MR imaging (Magnetic resonance imaging) has emerged as a key factor for the diagnosis of cancer.(11-18) Since it has advantages over noninvasive, good anatomical image due to high resolution,

high contrast and 3-dimensional information in real time more than nuclear medicine (PET, SPECT), optical imaging compared to other imaging modality.(19-23) And also, molecular MR imaging is able to detect simultaneously metabolism of cells and tissues and its physiological information and structural information, noninvasive and biological processes occurring in the deep tissues to provide the quantitative information.(24,25) Molecular MR imaging can observe a variety of imaging lesions as multi-modality in the diagnosis of breast cancer.

Many MR contrast agents have been used for good quality imaging (32-37). However passive contrast agents are not enough to reach their target goals specifically. Thus, we are aiming to develop intelligent targetable contrast agent using Hyaluronic acid (HA)(38-44). In particular, HA has become known to interact on CD44 receptor. Then breast cancer is known to being overexpressed CD44 receptor which as marker of cancer stem cell.(44) It is so crucial for MR probe to early breast cancer diagnostic point of view. Because CD44, important as cancer stem cell marker, is interacted with Hyaluronic acid. Hyaluronic acid which is a linear hydrogel with negative charge containing two alternating units of D-glucuronic acid (GLcUA) and N-acetyl-D-glucosamine (GLcNAc) with molecular weight of 105–107. HA has frequently been used for medical purposes such as a viscoelastic biomaterial in surgery.

Especially, it is well known that various human tumor cells (breast, ovarian, colon, lung, stomach, etc.) over-express HA-binding receptors, CD44 (44) In this study, molecular MR imaging were investigated to find biological processes which occur in gastric cancer. T2 weight sequence was simultaneously used to confirm for better diagnostic possibility and targeting effect was demonstrated through Hyaluronic acid conjugated magnetic nanoparticles in heterotopic xenograft breast cancer model. And various experiments were conduct to evaluate specific binding affinity and diagnostic effectiveness through in vivo and in vitro.

2. RESULT AND DISCUSSION

2.1 Preparation of MNPs and HA-MNPs

Monodispersed magnetic nanoparticles were synthesized using thermal decomposition and solubilized in nonpolar organic solvent, as previously reported. And HA conjugated MNP was synthesized using EDC and sulfo-NHS method. As shown in (Fig. 1), the characteristic band of HA-MNPs conjugates were verified by FT-IR spectra, which exhibits O-H stretching at 3200-3400 cm^{-1} , C=O stretching at 1100-1300 cm^{-1} , CO-NH(amide) bonds at 1630-1680 cm^{-1} and CH₂ bending in HA at 1430-1470 cm^{-1} .

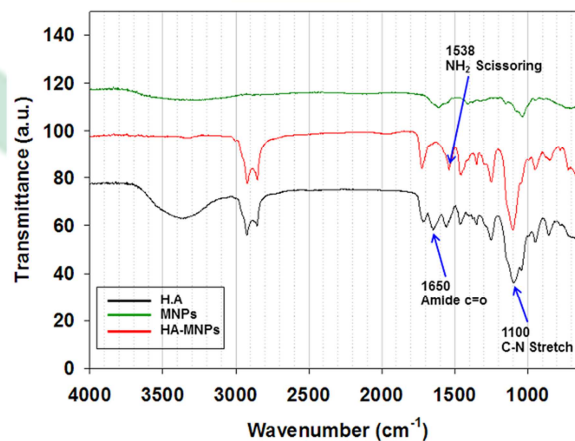


Fig.1. Fourier transform infrared spectra of HA (green line), MNP (red line) and HA-MNP(black line), i:CO-NH(amide) bonds

As MR agents, uniform MNPs (12nm) were synthesized at a high temperature via a thermal decomposition process. The size distribution and morphology of MNPs were

confirmed by transmission electron microscopy (Fig. 2.) which showed no significant differences in size or morphology between HA-MNPs and MNP. The size of the

water-soluble MNPs and HA-MNPs were determined to be $84.6 \pm 32.4\text{nm}$ and $137 \pm 53.2\text{nm}$, respectively. After the conjugation of Hyaluronic acid MNPs, the size slightly increased due to the large molecular weight of Hyaluronic

acid (1000kDa). In addition, the surface charge of aminated MNPs also changed from $20.62 \pm 1.96\text{ mV}$ (aminated water soluble MNP) to $-17.76 \pm 1.64\text{ mV}$ (HA coated MNP) due to the presence of HA (Fig. 2.).

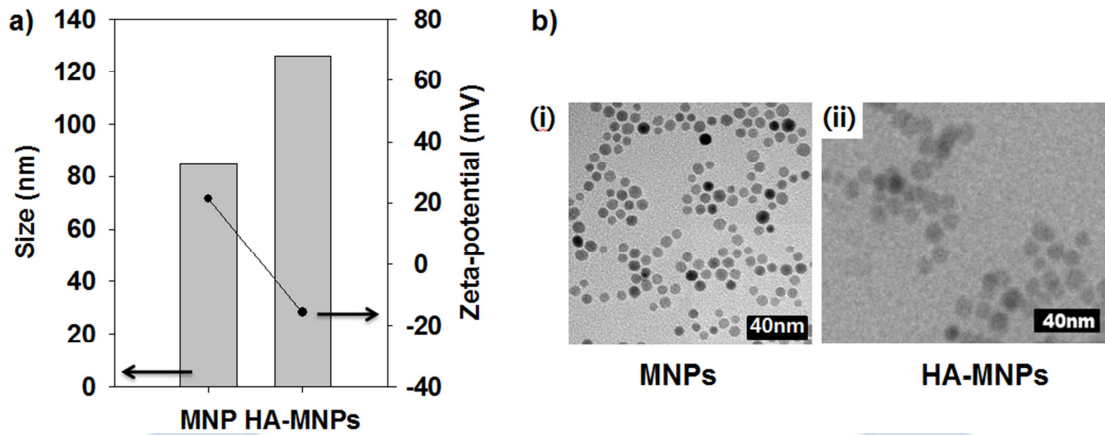


Fig.2. (a) is The average size (gray bar) and zeta potential (black circle) and (b) is TEM images of (i) magnetic nanoparticles (ii) hyaluronan-modified magnetic nanoparticles (HA-MNP)

2.2 Solubility and magnetic sensitivity of HA-MNPs

To assess the potential use of HA-MNPs as MR imaging agents, we performed MR imaging experiments using HA-MNPs, exhibiting the highest magnetic properties with appropriate size to avoid RES detection and prolong

retention in the circulation. In Fig. 2 (a) (i), the T2-weighted MR image exhibited a strong black color, which signified a decrease in signal intensity for the thicker HA-MNP solution. In Fig. 2 (b) (ii) represent change of intensity between of T2-weighted MR solution image

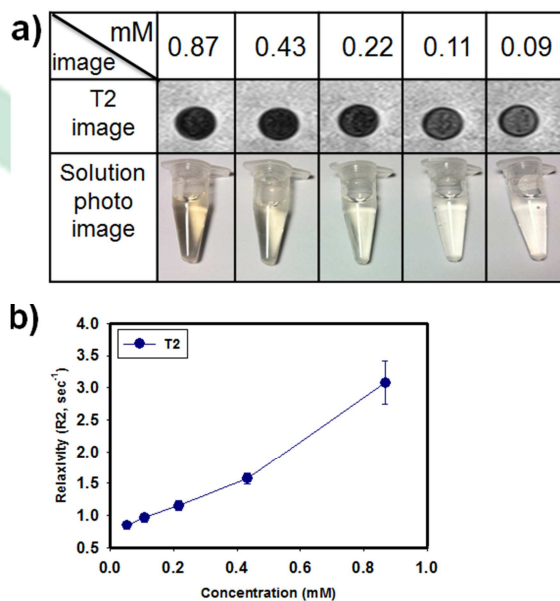


Fig.3. (a) Photographs and T2 solution MR images of HA-MNPs each conditions and (b) R2 relaxivity graph for the

magnetic ion concentration.

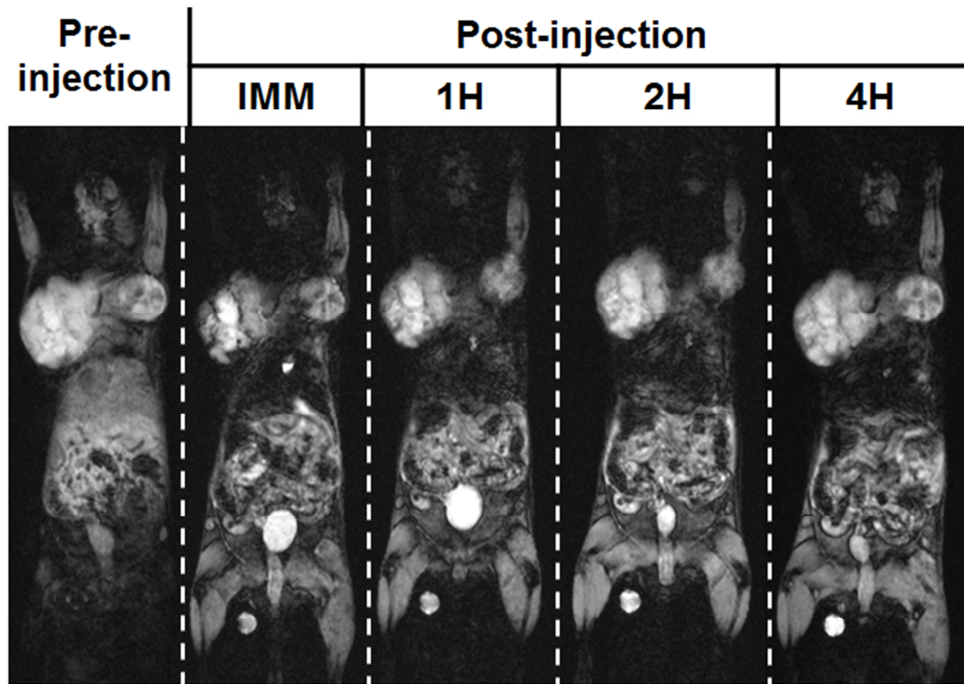


Fig.4. T2 MR images of tumor-bearing mice after intravenous injection of HA-MNPs. FOV: 100mm, ST: 1.00mm, TR: 14.85ms, TE: 5.65ms, coil elements: Wrist coil, Imaging modality: Siemens MR scanner

2.4 In vivo MR image

In Fig. 4, MR signal enhancement was identified after HA-MNPs injection. Initially, the center of tumor instantly darkened, and enhanced MR imaging signal intensity at surrounding vessels was simultaneously observed. In T2 weight MR images, clear anatomic details were observed,

3. CONCLUSION

In summary, we synthesized HA-MNPs as MR imaging agents for effective diagnosis for CD44-overexpressing breast cancer. Further study, we plan to evaluate as a molecular imaging tool using variable MR sequence for better imaging technique.

4. EXPERIMENTAL METHOD

4.1. Materials

Polysorbate 80, ethylenediamine, 1,4-dioxane (99.8%), 4-dimethylaminopyridine, triethylamine, and succinic anhydride (SA) were purchased from Sigma Aldrich Chemical Co. Phosphate buffered saline (PBS: 10 mM, pH 7.4), Roswell Park Memorial Institute-1640 (RPMI-1640), fetal bovine serum and antibiotic-antimycotic solution were purchased from Gibco and dialysis membrane (molecular weight cut off: 1,000 Da) was obtained from Spectrum laboratory. Hyaluronic acid (HA, 1,000,000 Da) was purchased from Yuhan Pharmaceutical Corporation (Seoul, Korea). MDA-MB-231 cell lines (American Type Culture Collection) were grown in medium containing 10% fetal bovine serum and 1% Antibiotic-Antimycotic at 37°C, humidified 5% CO₂ atmosphere.

4.2. Synthesis of magnetic nanoparticles

To synthesize monodispersed magnetic nanoparticles (MNPs), 2 mmol of iron (III) acetylacetonate, 1 mmol of manganese(II) acetylacetonate, 10 mmol of 1,2-hexadecanediol, 6 mmol of dodecanoic acid, and 6 mmol of dodecylamine were dissolved in 20 mL of benzyl ether under an ambient nitrogen atmosphere. The mixture was then pre-heated to 200°C for 2 hours and refluxed at 300°C for 30 minutes. After the reactants were cooled at room temperature, the products were purified with an excess of pure ethanol. Approximately 11 nm of MNPs were synthesized using the seed-mediated growth method.

4.3. Preparation of hyaluronan-modified magnetic nanoparticles (HA-MNPs)

HA-MNPs were prepared by the nano-emulsion method. 30 mg of magnetic nanoparticle were dissolved in 4 mL hexane (organic phase). This organic phase was poured into 20 mL deionized water (aqueous phase) containing 30 mg HA. The solution was ultra-sonicated in an ice-cooled bath for 20 min at 190W and stirred overnight at room temperature to evaporate the organic solvent. The resulting suspension was centrifuged three times for 20 min each at 3000 rpm. After the supernatant was removed, the precipitated HA-MNPs were re-dispersed in 5 mL deionized water. The size distribution and zeta potential of HA-MNPs were analyzed by laser scattering (ELS-Z, Otsuka Electronics); morphologies were confirmed using a transmittance electron microscope (TEM, JEM-2100, JEOL Ltd. Japan.). Finally, the relaxivity (R2) data of HA-MNPs solution were measured through magnetic resonance (MR) imaging analysis.

4.7. Heterotopic animal model and experimental procedure

All animal experiments were conducted with the approval of the Association for Assessment and Accreditation of Laboratory Animal Care (AAALAC) International. Female BALB/C-Slc nude mice at 7-8 weeks of age were anesthetized by intraperitoneal injection of a Zoletil/Rompun mixture and injected with 200 µL

containing 1.0×10^7 MDA-MB-231 cells suspended in saline into the femoral region. After cancer cell implantation, MR imaging was performed between 2 and 3 weeks. After MR imaging organ MR imaging performed too. In addition, the extracted tumor tissues from tumor-bearing mice treated with HA-MNPs were frozen, sectioned, and stained using Prussian blue. All stained tissue sections were analyzed using a virtual microscope (Olympus BX51, Japan) and Olyvia software.

4.8. MR imaging

Animal, organ, solution and cell MR imaging experiments were performed with a 3 Tesla Siemens clinical MRI instrument using a wrist coil with T2 Turbo spin echo sequence. (T2 Turbo spin echo: TR: 4000ms, TE : 114ms, Slice thickness : 1.0mm, FOV read : 180mm)

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